

- Background:** Initial studies by Thomas Arendt *et al.*, at the University of Leipzig identified the mitogenic response of peripheral blood lymphocytes (PLB) as a potential biomarker of Alzheimer's disease (AD) [Stieler, JT, and Arendt, T.; *et al.* Neuroreport 2001; 12(18):3969-3972]. This finding was confirmed in a second study conducted by Jens Steiler, *et al.*, where the authors were able to retrospectively differentiate AD from Other Dementias [(OD), mostly Idiopathic Parkinson's disease] with 95% sensitivity and 90% specificity [Steiler, J *et al.*, 2012. Neurobio Aging 33:234-341].
- Methods:** Analytical performance of this flow cytometer assay was re-established at a contract GLP laboratory. In collaboration with clinicians, patient blood was sampled and purified for PBMCs. Following the published protocol, a stimulation index was established for CD69 positive expression following mitogenic stimulation. Experiments were conducted on 12 healthy volunteer donor blood specimens to assess analytical performance characteristics of the assay. Pre-analytical and analytical variance was assessed in healthy normal subjects.
- Results:** Analytical performance data shows excellent reproducibility with coefficients of variation less than 20% for most mitogenic conditions. Additional performance metrics like effect of gating parameters and reagent stability were quantified. The effect of pre-analytical variables such as sample handling conditions were determined as well.
- Conclusion:** The LymPro test, which measures mitogenic response to peripheral blood lymphocytes, demonstrates suitable analytical performance for use in a fit-for-purpose fashion in the Company's hands

Review of Cell Cycle Dysregulation in Alzheimer's Disease

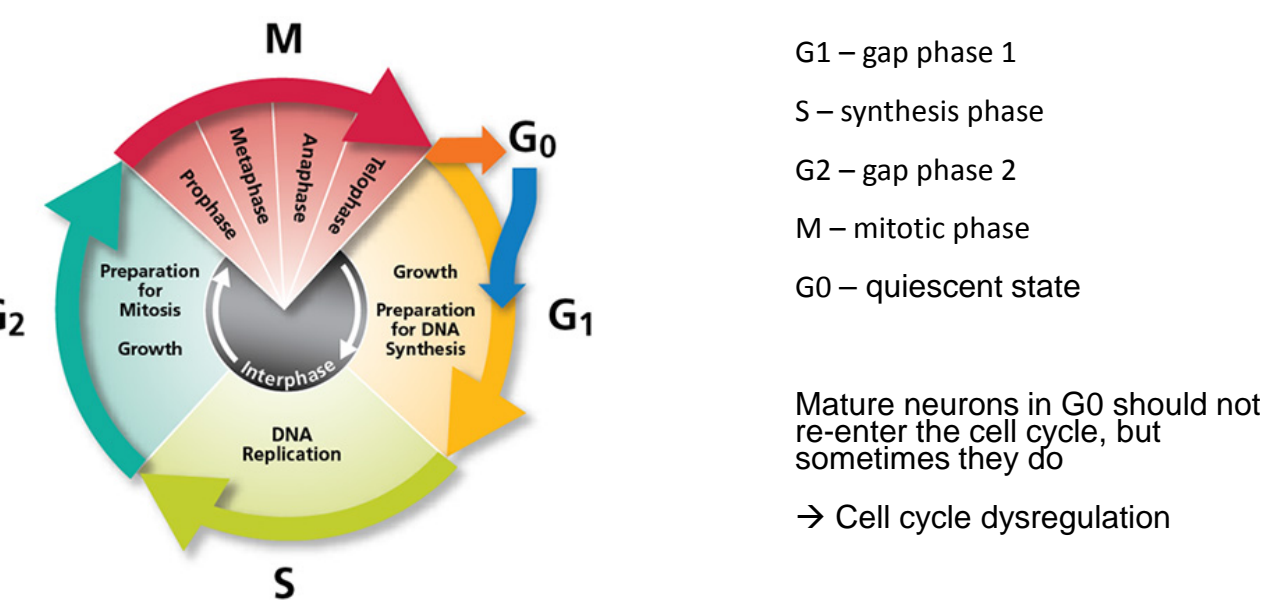


Figure 1

Table 1: Evidence for initiation of the cell cycle in neurons. Alzheimer's Disease (AD) patients show signs of neurons re-entering the cell cycle

Cell Cycle Proteins	Function	AD
Cyclin D	Important in G1 and for G1/S transition	↑
Cyclin E; CDK4; p21	Important in G1 and for G1/S transition	↑
PCNA	Facilitates DNA replication	↑
Cyclin G1	Important for G1/S transition	↑
Cyclin A	Involved in DNA replication and mitosis	↑
Cyclin B	Required for G2/M transition	↑
CDC2/CDK1	Regulates G1 progress and G1/S transition, promotes G2/M transition	↑
Ki67	Cellular proliferation marker present in all active phases (G1, S, G2, M), but not in resting cells (G0)	↑
P16	Inhibits cyclin D, CDK4/6 and G1 progression	↑

Table 2: Neuronal cell cycle re-entry is also apparent in mild cognitive impairment (MCI)

Cell Cycle Proteins	Function	MCI	AD
Cyclin D	Important in G1 and for G1/S transition	↑	↑
Cyclin B	Required for G2/M transition	↑	↑
Cyclin G1	Regulates G1/S transition	↑	↑
CDK2	Regulates G1/S transition	↑	
PCNA	Facilitates DNA replication	↑	↑

Table 3: Evidence for Cell Cycle Dysregulation in Lymphocytes as Surrogate for Neurons

Cell cycle protein / other	Biological role	AD
P53 mutant-like conformation	Regulation of G1/S cell cycle arrest; mutant conformation is functionally inactive	↑
Calmodulin	Regulation of G1/S cell cycle arrest	↑
Cyclin E	Regulation of G1 and for G1/S transition	↑
E ₂ F-1; CDK2; Rb (retinoblastoma)	Regulation of G1/S transition	↑
CD69	Lymphocyte development and migration	↓
Sensitivity to rapamycin and H ₂ O ₂	Blocks G1/S transition in healthy lymphocytes	↓

Possible links between neuron and lymphocyte cell dysregulation

- Production of Amyloid β (Aβ) Peptides in Alzheimer's disease exceeds the removal capacity of the brain
→ In brain, Aβ oligomers induce cell cycle re-entry in neurons
- Excess Aβ in body fluids activates peripheral lymphocytes and may lead to persistent stimulation of the immune system
→ May lead to disease-related alteration of lymphocytes
- Alteration may include dysregulation of CD69 gene expression regulation
→ Would affect lymphocyte proliferation and immune response in AD

Amarantus investigated an improved version of the LymPro assay

- Eight color antibody cocktail
- Becton Dickinson FACSCanto II flow cytometer

Antibodies and Fluorochromes

Antibody Target	CD3	CD69	CD14	CD4	CD28	CD45	CD19	CD8
Fluorochrome	FITC	PE	PERCP-CY5.5	PE-CY7	APC	APC-H7	V450	V500

Performance Characteristics

Table 4: CVs of CD69 measurements as a percent of lymphocyte class for various stimulation conditions.

Analyte	Stimulation	Median % CV	Range of %CV	Upper 95% CI Limit (% CV)	% of observations with > 20% CV
Total CD69* (as a % of CD45+ lymphs)	Unstim	7.5	0.6-15.9	10.9	0.0
Total CD69* (as a % of CD3*)	Unstim	8.2	0.0-29.9	20.3	22.2
Total CD69* (as a % of CD3*4*)	Unstim	27.9	0.0-56.8	39.6	77.8
Total CD69* (as a % of CD3*8*)	Unstim	10.3	5.0-20.6	16.6	11.1
Total CD69* (as a % of CD19*)	Unstim	5.3	0.3-8.9	7.2	0.0
Total CD69* (as a % of CD14*)	Unstim	18.8	4.4-81.3	44.3	44.4
Total CD69* (as a % of CD45+ lymphs)	PWM	5.8	0.3-9.9	7.9	0.0
Total CD69* (as a % of CD3*)	PWM	5.4	0.2-11.6	9.2	0.0
Total CD69* (as a % of CD3*4*)	PWM	4.7	0.2-11.8	8.9	0.0
Total CD69* (as a % of CD3*8*)	PWM	8.0	0.5-43.4	20.8	11.1
Total CD69* (as a % of CD19*)	PWM	4.9	0.3-6.9	6.0	0.0
Total CD69* (as a % of CD14*)	PWM	11.0	2.7-33.8	19.7	11.1
Total CD69* (as a % of CD45+ lymphs)	PHA	2.3	0.4-4.9	3.5	0.0
Total CD69* (as a % of CD3*)	PHA	2.2	0.4-5.1	3.5	0.0
Total CD69* (as a % of CD3*4*)	PHA	3.0	0.8-4.6	3.6	0.0
Total CD69* (as a % of CD3*8*)	PHA	2.1	1.0-4.7	3.5	0.0
Total CD69* (as a % of CD19*)	PHA	2.3	0.4-5.2	3.3	0.0
Total CD69* (as a % of CD14*)	PHA	9.0	2.4-20.1	12.9	11.1

PWM: Pokeweed mitogen; PHA: Phytohemagglutinin

Table 5: Summary of ANOVA for difference between fresh and 24 hr harvesting of PBMCs

Analyte	ANOVA p-value for CV difference between Fresh versus 24 hr harvest	ANOVA p-value for Mean difference between Fresh versus 24 hr harvest
Total CD69* (as a % of CD45+ lymphs)	0.7049	0.5690
Total CD69* (as a % of CD3*)	0.7687	0.5196
Total CD69* (as a % of CD3*4*)	0.1547	0.9577
Total CD69* (as a % of CD3*8*)	0.3196	0.4715
Total CD69* (as a % of CD19*)	0.0221	0.2779
Total CD69* (as a % of CD14*)	0.7180	0.2939

Table 6: General linear mixed models (GLMs) for CV

Analyte	p value, Donor	p value Stimulation	p value Fresh vs. 24 hr harvest
Total CD69* (as a % of CD45+ lymphs)	0.8354	0.0395	Nested within Donor
Total CD69* (as a % of CD3*)	0.6579	0.0207	Nested within Donor
Total CD69* (as a % of CD3*4*)	0.5704	<0.0001	Nested within Donor
Total CD69* (as a % of CD3*8*)	0.2032	0.0334	Nested within Donor
Total CD69* (as a % of CD19*)	0.0278	0.0156	Nested within Donor
Total CD69* (as a % of CD14*)	0.2043	0.0210	Nested within Donor

Table 7: GLMs for Mean value

Analyte	p value, Donor	p value Stimulation	p value Fresh vs. 24 hr harvest
Total CD69* (as a % of CD45+ lymphs)	0.0566	<0.0001	Nested within Donor
Total CD69* (as a % of CD3*)	0.0075	<0.0001	Nested within Donor
Total CD69* (as a % of CD3*4*)	0.0701	<0.0001	Nested within Donor
Total CD69* (as a % of CD3*8*)	0.0815	<0.0001	Nested within Donor
Total CD69* (as a % of CD19*)	0.0065	<0.0001	Nested within Donor
Total CD69* (as a % of CD14*)	0.0002	<0.0001	Nested within Donor

GLMs for CV and Mean value treating Donor and Stimulation condition as random effects, and Harvest point (Fresh or 24 hr) as a nested effect within Donor.

Summary

The Total CD69+ (as a % of CD45+ Lymphocytes) displays:

- Consistently low CVs in general and across stimulation conditions
- No effect of PBMC harvest point (Fresh or 24 hr)
- No inter-individual effect on CV in GLM models
- No inter-individual effect on Mean value in GLM models (which may indicate a tight distribution in the healthy control population)

The Total CD69+ (as a % of CD19+) also displays consistently low CVs as well as lack of significant difference on CV by stimulation condition, but shows an effect of harvest point and an inter-individual effect on CV.

Conclusions

The LymPro test, which measures mitogenic response to peripheral blood lymphocytes, demonstrates acceptable pre-analytical and analytical variation in this study. The test will transition into multiple clinical studies for differentiation of individuals at risk for Alzheimer's Disease from other dementias.

Key References

- Stieler, JT, and Arendt, T.; *et al.* Neuroreport 2001; 12(18):3969-39722.
- Stieler, J *et al.*, 2012. Neurobio Aging 33:234-341

For additional information:

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