

# #1716. Inhibition of c-Myc by APTO-253 as an Innovative Therapeutic Approach to Induce Cell Cycle Arrest and Apoptosis in Acute Myeloid Leukemia

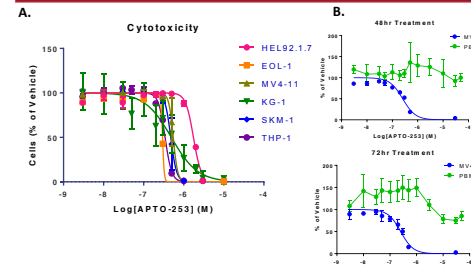
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## Abstract

c-Myc multifunctional transcription factor protein, a product on the c-myc proto-oncogene, contributes to the pathogenesis of many human cancers through multiple mechanisms of proliferation, apoptosis, cell cycle progression and senescence. c-Myc is frequently overexpressed in acute myeloid leukemia (AML), yet strategies to effectively and safely modulate c-Myc function do not exist. APTO-253 is a small molecule agent being developed clinically for the treatment of AML and high risk myelodysplastic syndromes (MDS), and we evaluated the effect of APTO-253 on c-myc gene expression. We first confirmed that c-Myc mRNA levels were significantly higher in AML cell lines as compared to peripheral blood mononuclear cells (PBMCs) isolated from healthy human donors. However, c-Myc expression in AML cells was inhibited by APTO-253 in dose-dependent and time-dependent manners at both the mRNA and protein levels, and c-Myc inhibition occurred as an early mechanistic event (6hr). Likewise, APTO-253 induced AML cell apoptosis in dose-dependent and time-dependent manners, as demonstrated by increases in Annexin-V staining and cleaved poly (ADP-ribose) polymerase (c-PARP). APTO-253 induced G1/G0 cell cycle arrest, increased p21 expression, decreased expression of cyclin D3 and cyclin-dependent kinases 4 (CDK4) in AML cells, and increased p53 levels in p53-positive MV4-11 and EOL1 cell lines. Collectively, these data suggest that inhibition of c-Myc by APTO-253 leads to global cell cycle arrest and apoptosis events in AML cells. Importantly, we demonstrated that APTO-253 selectively targeted tumor cells but not normal healthy cells, with MV4-11 AML cells and normal PBMCs having IC<sub>50</sub>s of 0.25±0.03μM and more than 100μM, respectively. Our previous studies (56<sup>th</sup> ASH abstract #4813) showed that APTO-253 induces the Krüppel-like Factor 4 (KLF4) transcription factor and was effective and well tolerated as a single agent in multiple AML xenograft models without causing bone marrow suppression. Taken together, our results indicate that nM levels of APTO-253 mechanistically inhibit c-Myc expression in AML cells and subsequently induce cell cycle arrest and apoptosis, and suggest that APTO-253 may serve as an effective and safe c-Myc inhibitor for AML.

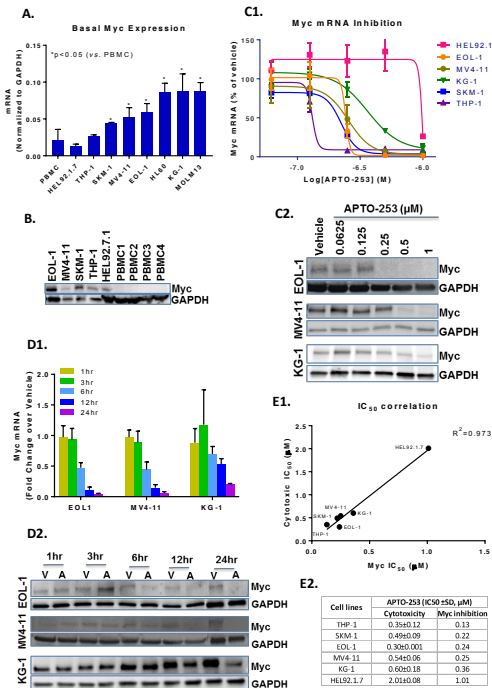
## APTO-253 Selectively Inhibits Proliferation of AML Cells But not Healthy Normal Cells



**Figure 1. Cytotoxic effect of APTO-253 on AML cells and healthy normal PBMCs.** Cells were plated and treated with and without APTO-253 at indicated concentrations in 96-well cell culture plates at 37°C. MTS based assay was performed at the end to quantify cell viability. Fifty percent cell growth inhibition (IC<sub>50</sub>) was calculated with GraphPad Prism 7. **A**, AML cells treated for 120 hours. Data from 3-7 independently experiments for different cell lines. **B**, PBMCs isolated from whole blood of 5 healthy human donors were treated for 48 and 72 hours in parallel with MV4-11 cells. MV4-11 cells and normal PBMCs had IC<sub>50</sub>s of 0.25±0.03μM and >100μM, respectively, at both time points.

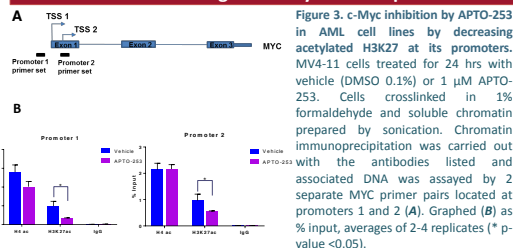
**Disclosures:** Zhang: Aptose Biosciences Inc.; Employment. Local: Aptose Biosciences Inc.; Employment. Benbatoul: Aptose Biosciences Inc.; Employment. Folger: Aptose Biosciences Inc.; Employment. Sheng: Aptose Biosciences Inc.; Employment. Esquivies: Aptose Biosciences Inc.; Employment. Lightfoot: Aptose Biosciences Inc.; Employment. Vellanki: Aptose Biosciences Inc.; Employment. Rice: Aptose Biosciences Inc.; Employment.

## APTO-253 Dose- and Time- Dependently Inhibits c-Myc in AML Cells



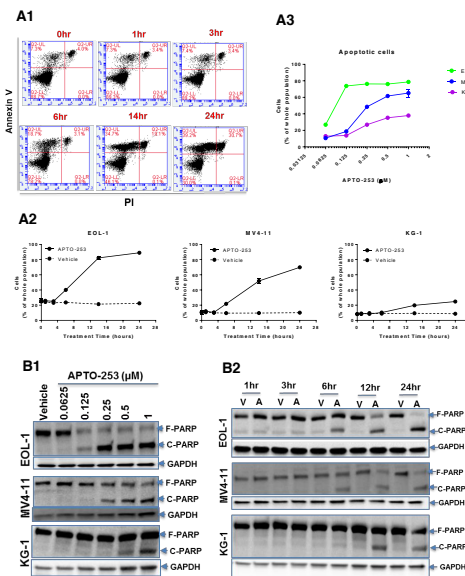
**Figure 2. c-Myc inhibition by APTO-253 in AML cell lines.** **A**, Basal Myc mRNA of AML cell lines and healthy normal PBMCs. RNA was extracted from non-treated cells and cDNA prepared from RNA. Myc expression was assayed by RT-qPCR and then normalized to GAPDH expression for each sample. Duplicate samples for HEL92.1.7, MOLM13, SKM1, and THP1, four replicates for EOL1 and KG1, six replicates for MV4-11 and nine replicates for healthy PBMCs. **B**, Basal Myc protein of AML cell lines and healthy normal PBMCs. The whole cell extract of non-treated cells was Western Blotted. **C**, Myc decreased by APTO-253 dose-dependently. Cells were treated with APTO-253 at indicated concentrations for 24 hours at 37°C. RNA (**C1**) and protein (**C2**, representative blots of at least 3 independent experiments) were extracted and quantified by RT-qPCR and Western Blot, respectively. Fifty percent Myc mRNA inhibition (IC<sub>50</sub>) was calculated with GraphPad Prism 7. **D**, Myc decreased by APTO-253 time-dependently. Cells were treated with APTO-253 at 0.5 μM for indicated time points at 37°C. RNA (**D1**) and protein (**D2**, representative blots of at least 3 independent experiments; v stands for vehicle treatment; A stands for APTO-253 treatment) were extracted and quantified by RT-qPCR and Western Blot, respectively. **E**, APTO-253 cytotoxic effect correlates with its Myc inhibitory effect. The correlation of IC<sub>50</sub>s of cytotoxicity (as described in Figure 1. A.) and Myc inhibition (as described in Figure 2. C.) was analyzed by GraphPad Prism 7.

## APTO-253 Deregulates Myc Transcription



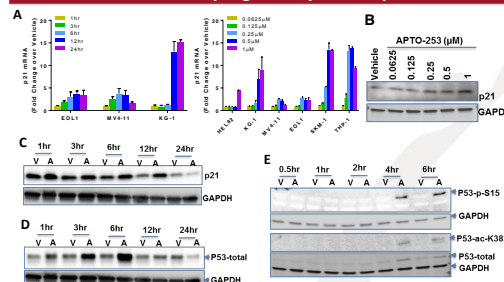
**Figure 3. c-Myc inhibition by APTO-253 in AML cell lines by decreasing acetylated H3K27 at its promoters.** MV4-11 cells treated for 24 hrs with vehicle (DMSO 0.1%) or 1 μM APTO-253. Cells crosslinked in 1% formaldehyde and soluble chromatin prepared by sonication. Chromatin immunoprecipitation was carried out with the antibodies listed and associated DNA was assayed by 2 separate MYC primer pairs located at promoters 1 and 2 (**A**). Graphed (**B**) as % input, averages of 2-4 replicates (\* p-value <0.05).

## APTO-253 Induces Apoptosis in AML Cells



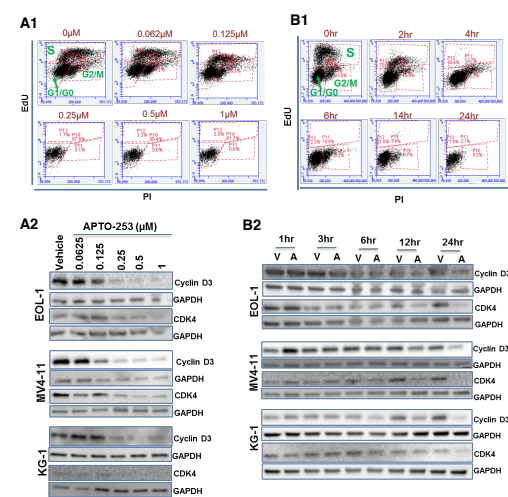
**Figure 4. APTO-253 dose-dependently and time-dependently induces apoptosis in AML cells.** AML cells EOL-1, MV4-11 and KG-1 were treated with vehicle DMSO (v) and APTO-253 (**A**) and subjected to apoptosis analysis by flow cytometry (**A1, A2 and B3**; **A1** as representative graphs of three independent experiments of three cell lines) and to apoptotic biomarker analysis by Western Blotting (**B1 and B2**, representative blots of three independent experiments for each cell line). For time course studies, cells were treated with 0.5μM APTO-253 for indicated times; for dose response studies, cells were treated with indicated concentrations for 24 hours. For the apoptosis analysis by flow cytometry, cells were stained with Annexin V and propidium iodide (PI) to distinguish live (Annexin V-/PI-), early apoptotic (Annexin V+/PI-) and dead (Annexin V+/PI+) cells. As indicated in **A2** and **A3**, total apoptotic cells (Annexin V+/PI- plus Annexin V+/PI+) were induced by APTO-253 in dose- and time- dependent manners, which agreed with PARP cleavage.

## APTO-253 Upregulates p21 and p53



**Figure 5. APTO-253 dose-dependently and time-dependently upregulates p21 and increases p53 activity in AML cells.** AML cells were treated with vehicle DMSO (v) and APTO-253 (**A**) at indicated concentrations for indicated times. The time course studies were done with 0.5μM treatment, and dose response studies were treated for 24 hours. The p21 mRNA (**A**) and p53 (**D**, **E**) Western Blots were shown for MV4-11 cells and represent three independent experiments. APTO-253 increases p21 expression at both mRNA and protein level. It increases total amount of p53 and its phosphorylation at Serine 15 and acetylation at Lysine 382 as early as 3-4 hour treatment

## APTO-253 Induces G1/G0 Cell Cycle Arrest in AML Cells



**Figure 6. APTO-253 dose-dependently and time-dependently induces cell cycle arrest in AML cells.** AML cells EOL-1, MV4-11 and KG-1 were treated with vehicle DMSO (v) and APTO-253 (**A**) and subjected to cell cycle analysis by flow cytometry (**A1 and B1**, representative graphs of three independent experiments of three cell lines) and to cell cycle biomarker analysis by Western Blotting (**A2 and B2**, representative blots of four independent experiments for each cell line). For time course studies, cells were treated with 0.5μM APTO-253 for indicated times; for dose response studies, cells were treated with indicated concentrations for 24 hours. For the cell cycle analysis by flow cytometry, cells were stained with 5-ethynyl-2 deoxyuridine (Edu) and propidium iodide (PI) to distinguish G1/G0, S and G2/M phases of cell cycle. Cell cycle biomarkers cyclin D1, cyclin D3, cyclin E, CDK4, CDK6, and CDK2 were Western Blotted. Only Cyclin D3 and CDK4 were consistently decreased by APTO-253 dose- and time-dependently in all three tested cell lines, which suggests cell cycle arrest at G1/G0 phase and is confirmed by flow cytometry analysis.

## Summary

- APTO-253 selectively kills AML cells without affecting healthy normal cells.
- APTO-253 inhibits c-Myc expression at both mRNA and protein levels in dose- and time- dependent manners. Its inhibitory effect on c-Myc correlates closely with its cytotoxic effect on AML cells (R<sup>2</sup>= 0.9731). Loss of acetylated H3K27 at Myc promoters suggests that APTO-253 transcriptionally deregulates c-Myc expression.
- APTO-253 dose- and time- dependently induces apoptosis and causes G1/G0 cell cycle arrest in AML cells.
- APTO-253 upregulates p21 and p53 at early time points (before 24 hour), which could contribute to apoptosis and cell cycle arrest.
- APTO-253 increases p53 stability and activity through inducing phosphorylation and acetylation, which could contribute to its mechanism of action in AML cells.
- Taken together, APTO-253 selectively targets tumor cells and effectively inhibits c-Myc expression in AML cells to induce cell cycle arrest and subsequent cell death.
- APTO-253 may serve as a safe and effective c-Myc inhibitor for AML chemotherapy that does not negatively impact the normal bone marrow cells.

## References:

1. Ito A, et al., EMBO. 2001. 20(6):1331-1340.
2. Shieh S, et al., Cell. 1997. 91:325-334.
3. Cercek A, et al., Invest New Drugs. 2015 Oct;33(5):1086-92